

THERAPEUTIC HYPOTHERMIA

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Living organisms according to the regulation of their body temperature are divided into two main groups, homoiotherms and poikilotherms. Homoiotherms are able to maintain constant their body temperature inspite of substantial changes of the environmental temperature. This is connected with all the time engagement of body metabolism and specific regulatory mechanisms. The body temperature of poikilotherms is closely dependent on the ambient temperature. In very low temperatures especially during winter the animals can survive by multiday hibernation which is realized by drop of their body temperature and torpor. The nature thus enables survival by means of pronounced reduction of energy expenditure, water loss and other physiological functions. In daily poikolitherms hypothermia and torpor lasting for several hours is used by diverse species in all climate zones.

Animals regulate their body temperature by means of hypothalamic thermoregulatory centre which coordinates the production and the release of the heat. Subcutaneous and mucosal thermoreceptors mediate peripheral thermoinformation. Heat production is influenced by metabolic processes which are hormonally modified and by shivering. In newborns nonshivering thermogenesis of the brown fat is used. Important role play vasomotoric reactins as vasoconstriction or vasodilatation. Mechanisms by which the organism gains or loosis temperature are dependent on the environmental temperature. Heat loss occurs via following mechanisms: radiation-emitation of heat in temperature lower than 37 °C, conduction – loss or gain of heat by the contact with cold or warm subjects inclusive of water, convection – due to the flow of air or water and evaporation – release of heat from the skin and the respiratory system (3, 15, 38, 66). The much higher thermal and ischemic tolerance of tissues of hibernators than those of homoiotherms led to the meditation about the use of hypothermia in medicine. Many animal and human studies setteld now recognized term therapeutic hypothermia, with its positive effects on metabolic, neurologic, cardiovascular and respiratory disturbances which are summarized in contemporary reviews (49, 55, 68).

At present there are many currently avaiable methods and devices for inducing and maintaining hypothermia (32, 41, 44). Principally they are divided to surface and core cooling. Surface cooling is realized by cold air exposure of skin with fans, air-circulating cooling blankets, fluid ice packs, prerefrigerated cooling pads, water-circulating cooling

blankets, selective head cooling etc. Core cooling is performed by intravenous catheters, infusion of ice-cold fluids, extracorporeal circulation, peritoneal lavage (tested in animals) antipyretic agents (with low efficacy). According to the dept following ranges of hypothermia are used: mild 35.9–34 °C, moderate 33.9–32 °C, moderate deep 31.9–30 °C and deep under 30 °C.

As to the course important is the speed of induction of hypothermia with better results when cooling is initiated rapidly. Duration of cooling depends on the severity of injury. Speed of rewarming should be slow with proper management and prevention of side effects (39). Induction of hypothermia is easier in old patients due to a slower counter-regulatory response and worse in obese patients due to the insulating properties of fat tissue. Eventual initial complication as fever can be suppressed by medication as well as shivering where skin counterwarming is also possible (12, 13, 14, 67). The key beneficial of therapeutic hypothermia is the reduction of the whole body metabolism. Basal and cerebral metabolic rates decrease by about 7% for every 1 °C reduction in core temperature. Decreased utilization of glucose, oxygen and high-energy phosphates mitigates secondary energy failure (61). Insulin release and secretion are decreased, glucagon secretion increases (10). Processes that are induced by anaerobic respiration are all ameliorated. During cooling are altered pharmacokinetics and pharmacodynamic of drugs (60).

Cardiovascular effects appear as decrease of heart rate, cardiac output and arterial pressure (35). Heart rate is reduced by about 10 °C reduction in body core temperature. Also cardiac output decreases linearly with variable effects on stroke volume. Slowing diastolic repolarization increases both myocardial conduction time and absolute refractory period concomitantly with the depression of sympathetic autonomic nervous system (19). ECG changes include prolonged PR, QRS and QT intervals and in adults positive deflection between the QRS and ST segments- Osborne or J wave (12). The decrease of cardiac output is compensated by total peripheral resistance. This and the hypoxia induced loss of cerebral autoregulation maintains the cerebral perfusion. Mild hyperthermia may increase membrane stability, thereby decreasing the risk of arrhythmias in contrast to deep hypothermia (40).

Hypothermia suppresses nonconvulsive and convulsive status epilepticus as well as spreading depression. There is a linear relationship between body temperature and EEG voltage. Deep hypothermia below 23 °C results in an isoelectric EEG. Nevertheless during this condition it is possible in rats and dogs to provoke epileptogenic discharges after local application of strychnine and acetylcholine to the brain cortex. Thus EEG is not a reliable indicator in the prognosis of outcome of brain ischemia (54). Hypothermia inhibits harmful excitatory processes occurring in brain cells during ischemia reperfusion, suppresses inflammatory reactions and release of inflammatory cytokines, reduces generation of free radicals, decreases intracranial pressure which ameliorates cytotoxic edema (7).

Hypothermia may affect a broad spectrum of haematological parameters as coagulation and leucocyte function with increased risk of bleeding and infection (4, 22, 28, 30, 43, 65). Therefore application of antibiotics is recommended. Useful is also decontamination of the digestive tract (11). However in asphyxiated children prolonged hypoxia may have already altered the haemostasis before cooling is initiated.

Hypothermia may provide beneficial effects for the abdominal viscera and is hepato – and nephroprotective. It does not increase the risk of necrotising enterocolitis and reduces ischemic – reperfusion injury and microvascular permeability. During cooling it is preferable to withheld enteral feeding to prevent additional stress on intestines.

Prerequisite for successful application of hypothermia in the clinical praxis were animal experiments. Many animal species were used – rats, rabbits, cats, dogs, piglets, apes (24, 25, 32, 42, 45, 57). Dogs survived asystole after cooling to 1.5 °C, the heart beating again upon rewarming the blood without any clinically detectable sequelae during a one month observation period (64). Numerous are experiments with tolerance to anoxic ischemia in newborn and adult animal models (1, 16, 18, 24, 25, 31). In newborn dogs during profound hypothermia cerebral glucose utilisation was globally depressed but cerebral blood flow remained more than adequate to support the energy needs of the brain (17). Brain hypothermia protects against ischemic neuronal damage even in the aged animals. Hypothermia may prevent cerebral edema and neuropathologic damage associated with hypoxic-ischemic injury in neonatal rats. Hypothermia after severe transient hypoxia delayed energy failure in newborn piglets and protected the spinal cord from ischemic injury in a chronic porcine model (58, 59). Hypothermic preconditioning increased survival of purkinje cells in rat cerebellar slices after an *in vitro* simulated ischemia (69). But there were described detrimental effects of very prolonged hypothermia in cats and monkeys with and without regional cerebral ischemia. Also in other experiments with rats deep prolonged hypothermia was followed by derangement of peripheral circulation (56).

In clinical disciplines hypothermia is mainly used in internal medicine dealing with acute heart failure, in surgery dealing with complicated operations, in neurology dealing with brain apoplexy, in neonatology dealing with anoxic-ischemic encephalopathy and in reanimatology. Studies with cardiac arrest noted significant improvement with hypothermia implementation (2, 6, 23, 48, 52, 72). It is recommended to cool most cardiac arrest to 32 °C with optimal duration of 24, 48 or 72 hours always with a slow rewarming rate. Questionable is the benefit of hypothermia in prehospital patients with out-of-hospital cardiac arrest (27).

Even patients with nonshockable rhythms should have a chance of full recovery (48). Antegrade and retrograde cerebral perfusion combined with deep hypothermia has protective effect in operations with circulatory arrest (26).

Hypothermia has neuroprotective effect after cardiac arrest (20, 34, 68, 69). Focal cooling suppresses spontaneous epileptiform activity without changing the cortical motor threshold (29), can be used for refractory status epilepticus (9) and ameliorates epileptic brain damage, while hyperthermia aggravates it (8, 21b, 33, 37, 53).

Hypothermia is a potential neuroprotective intervention to treat neonatal hypoxic-ischemic encephalopathy. Meta analysis of 13 clinical trials proved reproducible reduction of mortality, neurodevelopmental disability, cerebral palsy, cognitive and psychomotor delay (49). Other publications are more critical (5, 50, 51).

As an extensive independent sphere is the use of hypothermia in cryosurgery (21a). Automated apparatuses and diverse instruments cooled by liquid nitrogen (–196 °C) and

nitrous oxide ($-89.5\text{ }^{\circ}\text{C}$) are used. Surgical interventions are possible in every tissue and organ inclusive of tumours.

Quite new is cryotherapy using for cooling of the whole body extremely low temperature of $110\text{--}160\text{ }^{\circ}\text{C}$. In the Czech Republic it was introduced in the year 2004 in Čeladná. The procedure in this equipment passes in two steps. Clothed person stays a short time in ante-room with temperature $-60\text{ }^{\circ}\text{C}$. After crossing into the main chambre with temperature $-120\text{--}130\text{ }^{\circ}\text{C}$ the stay is limited by 3 minutes. The result is a rapid cooling of the body surface with great hyperemia lasting for several hours. The described positive health effects are complex.

SUMMARY

In a review there are described the mechanisms of thermoregulation in homoio – and poikilotherms, the use of hypothermia in animal experiments, in the clinical praxis, in cryosurgery and in cryotherapy.

Terapeutická hypotermie

SOUHRN

V přehledu jsou popsány mechanismy termoregulace u poikilo- a homiotermů, využití hypotermie v pokusech na zvířatech, v klinické praxi, v kryochirurgii a v kryoterapii.

REFERENCES

1. Alva N., Azura D., Palomague J. et al.: Deep hypothermia protects against acute hypoxia in vivo in rats: a mechanism related to the attenuation of oxidative stress. *Exp. Physiol.* 98, 2013: 1115–24. – 2. Arrich J., Holzer M., Havel C.: Hypothermia for neuroprotection in adults after cardiopulmonary resuscitation. *Cochrane database Syst. Rev.* 2009. – 3. Azzopardi D., Edwards A. D.: Hypotherm. *Semin. Fetal Neonatal Med.* 12, 2007: 303–10. – 4. Bacher A.: Effects of temperature on blood gases. *Intensiv. Care Med.* 31, 2005: 11–24. – 5. Blackmon L. R., Stark A. R.: Hypothermia: A neuroprotective therapy for neonatal hypoxia-ischemic encephalopathy. *Pediatrics* 117, 2006: 942–8. – 6. Boddecker K. A., Khang Y., Zimmerman M. B.: Hypothermia improves defibrillation success and resuscitation outcomes from ventricular fibrillation. *Circul.* 111, 2005: 3195–201. – 7. Busto R., Globus M. Y., Dietrich W. B. et al.: Effect of mild hypothermia on ischemia induced release of neurotransmitters and free fatty acids in rat brain. *Stroke* 20, 1989: 904–10. – 8. Commichau C., Scarmeas M. A., Segal A. Z. et al.: Risk factors for fever in the neurologic intensive care unit. *Neurology* 60, 2003: 837–41. – 9. Corry J. J., Dhar R., Murphy T. et al.: Hypothermia for refractory status epilepticus. *Neurocrit. Care* 9, 2008: 189–97. – 10. Cueni-Viloz N., Devigili A., Delodder F. et al.: Increased blood glucose variability during hypothermia and outcome after cardiac arrest. *Crit. Care Med.* 39, 2011: 2225–31. – 11. De Jonge E., Schultz M. J., Sponjaar J. et al.: Effects of selective decontamination of digestive tract on mortality and acquisition of resistant bacteria in intensive care: A randomised controlled trial. *Lancet* 362, 2003: 1011–6. – 12. De Witte J., Sessle D. I.: Perioperative shivering: Physiology and pharmacology. *Anesthesiol.* 96, 2002: 467–84. – 13. Diaz M., Becker D. E.: Thermoregulation: physiological and clinical consideration during sedation and general anesthesia. *Anest. Progr.* 57, 2010:

25–33. – 14. Diringer M. N., Reaven N. L., Funk S. E. et al.: Elevated body temperature independently contributes to increased length of stay in neurologic intensive care patients. *Crit. Care Med.* 32, 2004: 489–95. – 15. Doherty A. H., Florant G. L., Donahue S. W.: Endocrine regulation of bone and energy metabolism in hibernating mammals. *Integr. Comp. Biol.* 54, 2014: 463–83. – 16. Ehrlich M. P., Mc Cullough J. N., Zhang N. et al.: Effect of hypothermia on cerebral blood flow and metabolism in the pig. *Ann. Thor. Surg.* 73, 2002: 191–7. – 17. Emsie-Smith D., Sladden G. E., Stirling G. R.: The significance of changes in the electrocardiogram in hypothermia. *Br. Heart J.* 21, 1959: 343–51. – 18. Erecinska M., Thoresen M., Silver I. A. et al.: Effects of hypothermia on energy metabolism in mammalian central nervous system. *J. Cerebr. Blood Flow* 23, 2003: 513–30. – 19. Espinoza A., Keraus V., Opdahl A. et al.: Effects of therapeutic hypothermia on left ventricular function assessed by ultrasound imaging. *J. Am. Soc. Echocardiogr.* 11, 2013: 1252–63. – 20. Froehler M. T., Geocadin R. G.: Hypothermia for neuroprotection after cardiac arrest: mechanism, clinical trials and patient care. *J. Neurol. Sci.* 261, 2007: 118–26. – 21a. Gage A. A., Baust J. M., Baust J. G.: Experimental cryosurgery investigations in vivo. *Cryobiology* 59, 2009: 228–43. – 21b. Gasparini A., Guo Y., Hashizuma M. et al.: Mortality risk attributable to high and low ambient temperature: a multicountry observational study. *Lancet* 386, 2015: 369–75. – 22. Geurts M., Macleod M. R. et al.: Therapeutic hypothermia and the risk of infection: a systemic review and meta-analysis. *Crit. Care Med.* 42, 2014: 231–42. – 23. Gillmann H., Zindler M., Löhr B.: Über die Wirkung der tiefen künstlichen Hypothermie auf die Aktion des menschlichen Herzens. *Arch. Kreislaufforsch.* 27, 1957: 288–305. – 24. Goldberg L. I.: Effects of hypothermia on contractility of the intact dog heart. *Amer. J. Physiol.* 194, 1958: 92–8. – 25. Hagerdal M., Harp J., Nilsson L. et al.: The effect of induced hypothermia upon oxygen consumption in the rat brain. *J. Neurochem.* 24, 1975: 311–6. – 26a. Horvath S. M., Spurr G. B., Hutt B. K. et al.: Metabolic cost of shivering. *J. Appl. Physiol.* 8, 1956: 595–602. – 26b. Hu Z., Wang Z., Ren Z. et al.: Similar cerebral protective effectiveness of antegrade and retrograde cerebral perfusion combined with deep hypothermia circulatory arrest in aortic heart surgery: a meta-analysis and systematic review of 5060 patients. *J. Thorac. Cardiovasc. Surg.* 148, 2014: 544–60. – 27. Huang F. Y., Huang B. T., Wang P. J.: The efficacy and safety of prehospital therapeutic hypothermia in patients with out-of-hospital cardiac arrest: A systematic review and meta-analysis. *Resuscitation* 21, 2015: 170–9. – 28. Childs E. W., Udobi K. F., Hunter F. A.: Hypothermia reduces microvascular permeability reactive oxygen species expression after hemorrhagic shock. *J. Trauma* 58, 2005: 271–7. – 29. Karkar K. M., Garcia P. A., Bateman I. M.: Focal cooling suppresses spontaneous epileptiform activity without changing the cortical motor threshold. *Epilepsia* 43, 2002: 932–5. – 30. Kelman G. R., Nunn J. F.: Nomogram for correction of blood PO₂, PCO₂, pH and base excess and temperature. *J. Appl. Physiol.* 21, 1966: 1484–90. – 31. Kimura T., Sako K., Tamaka K. et al.: Effect of milde hypothermia on energy state recovery following transient forebrain ischemia in the gerbil. *Exp. Brain Res.* 145, 2002: 83–90. – 32. Knochner P.: Effects of experimental hypothermia on vital organs. *Lancet* 1955: 837–40. – 33. Kurz A.: Thermal care in the perioperative period. *Best. Pract. Res. Clin. Anesthesiol.* 22, 2008: 39–62. – 34. Lavinio A., Timofeev I., Nort J. et al.: Cerebrovascular reactivity during hypothermia and rewarming. *Br. J. Anaesth.* 99, 2009: 237–44. – 35. Lewis M. E., Al-Khalidi A. H., Blennov G. et al.: The effect of hypothermia on human left ventricular contractile function during cardiac energy. *J. Am. Coll. Cardiol.* 39, 2002: 102–8. – 36. Lu S. H., Leasure A. R., Dai Y. T.: A systematic review of body temperature variations in older people. *J. Clin. Nurs.* 19, 2010: 4–16. – 37. Lundgren J., Smith M. L., Blennov G.: Hyperthermia aggravates and hypothermia ameliorates epileptic brain damage. *Brain Res.* 99, 1994: 43–55. – 38. Malan A.: The evolution of mammalian hibernation: lessons from comparative acid-base physiology. *Integr. Comp. Biol.* 54, 2014: 96. – 39. Maxwell W. L., Watson A., Queen R. et al.: Slow, medium or fast re-warming following post-traumatic hypothermia therapy. An ultrastructure perspective. *J. Neurotrauma* 22, 2005: 873–84. – 40. Mirzoyev S. A., McLeod C. S., Burch T. J. et al.: Hypokalemia during the cooling phase of therapeutic hypothermia and its impact on arrhythmogenesis. *Resusc.* 81, 2010: 1632–6. – 41. Moore E. M., Nichol A. D., Bernard S. A. et al.: Therapeutic hypothermia: benefits, mechanisms and potential clinical applications in neurological, cardiac and kidney injury. *Injury* 42, 2011: 843–54. – 42. Morgan M. L., Anderson H. J., Ellis M. A. et al.: Mechanism of cold diuresis in the rat. *Am. J. Physiol.* 244, 1983: F210–6. – 43. Momvillier B., Tubach F., van de Beek D. et al.: Induced hypothermia in severe bacterial meningitis: a randomised clinical trial. *JAMA* 310, 2013: 2174–83. – 44. Nolan P., Soar J., Zideman D. A. et al.: ERG guidelines writing group. European resuscitation council guidelines for resuscitation. *Resusc.* 10, 2010: 1219–76. – 45. Oboshi H., Ibayashi S., Takano K. et al.: Hypothermia inhibits ischemia-induced efflux of amino acids and neuronal damage in the

hippocampus of aged rats. *Brain Res.* 884, 2000: 23–30. – 47. Palmer Ch., Vannucci R. C., Christensen M. A. et al.: Regional cerebral blood flow and glucose utilization during hypothermia in newborn dogs. *Anesthesiol.* 71, 1989: 730–7. – 48. Polderman K. H., Varon J.: How low should we go? Hypothermia or strict normothermia after cardiac arrest? *Circul.* 131, 2015: 669–75. – 49. Shah P. S.: Hypothermia: a systematic review and meta-analysis of clinical trials. *Seminars Fetal Neonatal Med.* 15, 2010: 238–240. – 50. Shankaran S., Laptook A. R., Pappas A. et al.: Effect of depth and duration of cooling on deaths in the NICU among neonates with hypoxic ischemic encephalopathy. A randomised clinical trial. *JAMA* 312, 2014: 2029–39. – 51. Shankaran S., Laptook A., Wright L. L. et al.: Whole-body hypothermia for neonatal encephalopathy: Animal observations as a basis for a randomized, controlled pilot study in term infants. *Pediatrics* 110, 2002: 377–81. – 52. Scheföld J. C., Storm C., Joerres A. et al.: Mild hypothermia after cardiac arrest and the risk of bleeding in patients with acute myocardial infarction. *Int. J. Cardiol.* 132, 2008: 387–91. – 53. Schwarz S., Hafner K., Aschoff A. et al.: Incidence and prognostic significance of fever following intracerebral hemorrhage. *Neurology* 54, 2000: 354–61. – 54. Sobotka P.: Isoelectric EEG and the function of the brain cortex. *Plzeň. lék. Sborn.* 70, 2003: 15–22. – 55. Sosnowski P., Mikrut K., Krauss H.: Hypothermia-mechanism of action and pathophysiological changes in the human body. *Postepy Hig. Med. Dosw.* 69, 2015: 69–79. – 56. Steen P. A., Soule E. H., Michenfelder J. D.: Detrimental effect of prolonged hypothermia in cats and monkeys with and without regional cerebral ischemia. *Stroke* 10, 1979: 522–9. – 57. Tveita T., Mortensen E., Hevroy O. et al.: Experimental hypothermia: Effects of core cooling and rewarming on hemodynamics, coronary blood flow, and myocardial metabolism in dogs. *Anesth. Analg.* 79, 1994: 212–8. – 58. Strauch J. P., Lauten A., Spielvogel D. et al.: Mild hypothermia protects the spinal cord from ischemic injury in a chronic porcine model. *Eur. J. Thorac. Surg.* 25, 2004: 708–15. – 59. Thoresen M., Penrice J., Lorek A. et al.: Mild hypothermia after severe transient hypoxia-ischemia ameliorates delayed energy failure in the newborn piglet. *Pediatr. Res.* 37, 1995: 667–70. – 60. Toriorici M. A., Kochauer P. M., Poloyac S. M.: Effects of hypothermia on drug disposition metabolism and response: A focus of hypothermia-mediated alterations on the cytochroma P 450 enzyme system. *Crit. Care Med.* 35, 2007: 2196–204. – 61. Tran C., Gariani K., Herrmann F. R. et al.: Hypothermia is a frequent signe of severe hypoglycaemia in patients with diabetes. *Diab. Metab.* 38, 2012: 170–3. – 62. Tveita T., Ytrehus K., Myhre S. et al.: Left ventricular dysfunction following rewarming from experimental hypothermia. *J. Appl. Physiol.* 85, 1998: 2135–9. – 63. Tveita T., Ytrehus K., Skandfer M. et al.: Changes in blood flow distribution and capillary function after deep hypothermia in rat. *Can. J. Physiol. Pharmacol.* 74, 1996: 376–81. – 64. Tysinger D. S., Grace J. T., Gollan F.: The electrocardiogram of dogs surviving 1.6 centigrade. *Am. J. Physiol.* 174, 1995: 816–22. – 65. Valeri C. R., Feingold H., Cassidy A. et al.: Hypothermia-induced reversible platelet dysfunction. *Ann. Surg.* 205, 1987: 175–81. – 66. Van Breukelen F., Martin S. L.: The hibernation continuum: Physiological and molecular aspects of metabolic plasticity in mammals. *Physiology* 30, 2015: 273–81. – 67. Van Zante. A. R. H., Polderman K. H.: Skin countervarming to prevent shivering during therapeutic cooling. *Crit. Care Med.* 37, 2009: 2006–8. – 68. Weng Y., Sun S.: Therapeutic hypothermia after cardiac arrest in adults: mechanism of neuroprotection, phases of hypothermia and methods of cooling. – 69. Yuan H. B., Huang Y., Zhang S.: Hypothermic preconditioning increases survival of purkinje neurons in cat cerebellar slices after an *in vitro* simulated ischemia. – 70. Young S. K., Olinginski T. P., Yagel S. K. et al.: The effect of graded hypothermia on hypoxic-ischemic brain damage: A neuropathologic study in the neonatal rat. *Stroke* 14, 1983: 929–34. – 71. Yunoki M., Nishio S., Ukita N. et al.: Hypothermic preconditioning induces rapid tolerance to focal ischemia injury in the rat. *Exp. Neurol.* 181, 2003: 291–300. – 72. Zanella S., Buck M., Fairchild K.: Physiologic and pharmacologic consideration for hypothermia therapy in neonates. *J. Perinatol.* 31, 2011: 377–86. – 73. Zhao M., Shimohata T., Wang J. Q. et al.: Akt contributes to neuroprotection by hypothermia against cerebral ischemia in rats. *J. Neurosci.* 25, 2005: 9794–06.

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