

# Statins and Dental Implant Osseointegration – Bridging Molecular Science and Next Generation Clinical Outcomes: A Review

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## ABSTRACT

Statins, widely prescribed for dyslipidemia, may exert bone modifying effects through coordinated stimulation of osteoblast activity and suppression of osteoclastogenesis. Lipophilic statins, including simvastatin, lovastatin, and atorvastatin, have been shown in preclinical models to enhance osteoblast differentiation and matrix synthesis via BMP-2/Smad and Ras-PI3K-Akt-Erk signaling, while attenuating osteoclast development through modulation of the OPG/RANKL axis, NF-κB inhibition, and blockade of p38 MAPK pathways. Despite mechanistic consistency in experimental systems, human data remain inconclusive, with modest increases in bone mineral density and no confirmed reduction in fracture risk. In implant dentistry, hyperlipidemia has been linked to impaired osseointegration, likely via reduced osteoblastic function, increased osteoclast activity, and compromised collagen turnover. The interplay between obesity, lipid metabolism, and skeletal biology introduces additional confounding variables, emphasizing the need for patient stratification in clinical research. Current evidence supports the biological plausibility of statin mediated enhancement of peri-implant bone formation, but definitive clinical translation requires large-scale, stratified trials with controlled delivery approaches and extended follow-up.

## KEYWORDS

statins; dental implants; osseointegration; bone metabolism; osteoblasts and osteoclasts

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## INTRODUCTION

Beyond their lipid lowering properties, statins may influence skeletal biology through pleiotropic mechanisms that converge on bone forming and bone resorbing cells (1). The inhibition of HMG-CoA reductase reduces isoprenoid intermediates, impairing prenylation-dependent signaling in small GTPases such as Rho and Ras (2). This molecular interference appears to enhance osteoblastic activity and limit osteoclastic differentiation in experimental systems (1, 3, 4). Currently there is an academic interest in the potential of statins to improve bone mineral density (BMD) and support osseointegration, particularly under conditions of metabolic disturbance such as hyperlipidemia.

The clinical relevance of these effects remains uncertain, as epidemiological associations and modest trial results have yet to translate into unequivocal fracture risk reduction. Moreover, the interplay between obesity, dyslipidemia, and bone metabolism introduces additional variables that complicate interpretation. In dental and orthopedic contexts, the possibility of local statin delivery to enhance bone healing warrants investigation, especially for patients with compromised lipid profiles.

## STATINS AND OSTEOLASTS

Statins may stimulate osteoblasts through multiple molecular pathways. In cell cultures, simvastatin enhances alkaline phosphatase activity and mineralization in MC3T3-E1 osteoblastic cells, acting in a dose-dependent manner and raising BMP-2 and ALP mRNA expression, while suppressing collagenase-1 expression – a signature of anabolic activity (5). More detailed mechanistic work shows simvastatin promotes osteoblast differentiation via a membrane-bound Ras/Smad/Erk/BMP-2 pathway. Treatment increases ALP activity and upregulates genes for BMP-2, sialoprotein, and type I collagen (6). Across animal and in vitro models, statins, especially lipophilic ones like simvastatin, lovastatin, and atorvastatin, appear to modulate small GTPases (Ras, Rho), suppressing their prenylation and thus enabling osteogenic signaling, while also engaging TGF- $\beta$ /Smad3 pathways to reduce osteoblast apoptosis (7).

A broader review emphasizes these pleiotropic effects, noting that statins increase mediators such as BMP-2, TGF- $\beta$ , ALP, type I collagen, and collagenase-1 to bolster bone formation (8). These findings suggest that statins may support bone tissue by enhancing osteoblastic survival, differentiation, and matrix production, though the clinical margin of effect on bone healing or osteoporosis remains to be fully confirmed.

## STATINS AND OSTEOCLASTS

Statins appear to suppress osteoclast development and activity. Experimental studies report that statins increase osteoprotegerin (OPG), a decoy receptor that inhibits RANKL, and decrease RANKL levels, altering the OPG/

RANKL/RANK axis in favor of reduced osteoclastogenesis (9). In vitro, simvastatin blocks RANKL-induced transcriptional activation of NF- $\kappa$ B by preserving I $\kappa$ B $\alpha$ , suppressing osteoclast differentiation. In vivo, simvastatin reduces osteoclast numbers and IRF4 expression in rodent models, increasing bone volume and preventing bone loss when RANKL is experimentally elevated (10). Lovastatin likewise inhibits osteoclastogenesis in a dose-dependent fashion in rat bone marrow cultures.

## CORRELATION OF OBESITY TO HYPERCHOLESTEROLEMIA

Obesity and elevated cholesterol frequently coexist, though the causative threads are intricate. Excess adipose, especially visceral fat, releases adipokines and inflammatory signals that may boost hepatic lipid synthesis and compromise lipid clearance, potentially raising LDL cholesterol (11). Insulin resistance, common in obesity, may augment hepatic VLDL production and elevate circulating lipids. Epidemiological data frequently shows positive correlations between BMI or waist circumference and LDL or total cholesterol, though these correlations vary by age, diet, genetics, and ethnicity (12).

Obesity may act less as a direct driver and more as part of a metabolic profile that includes genetic predispositions, sedentary habits, dietary patterns, and, increasingly recognized, gut microbiome composition (13). Thus, while obesity increases the likelihood of hypercholesterolemia, particularly within metabolic syndrome, it is not sufficient to predict it. Instead, it contributes to a constellation of risk factors shaping lipid dysregulation.

## HYPERCHOLESTEROLEMIA AND OSSEOINTEGRATION OF DENTAL IMPLANTS

Hyperlipidemia seems to impair the healing processes underpinning osseointegration. In mice, a high fat diet reduced peri-implant bone volume, decreased mechanical strength at the bone to implant interface, and increased implant loss (1); these effects reflect reduced osteoblastic activity, enhanced osteoclast differentiation, disrupted collagen processing, and impaired bone quality (14). Osseointegration, the direct structural and functional connection between bone and implant surface, depends heavily on osteoblast mediated matrix deposition and remodeling. Hyperlipidemia appears to disrupt both of those key processes.

Statin delivery strategies, including local application via coatings, hydrogels, nanoparticles, and bone graft materials, have shown promise in promoting bone formation and improving osseointegration in hyperlipidemic contexts (4). For example, simvastatin coated on  $\beta$ -tricalcium phosphate or delivered via PLGA-PEG hydrogels can enhance peri-implant bone healing, though evidence is largely preclinical and further validation is required (7, 15).

Consequently, while hypercholesterolemia may pose a risk to implant success, adjunctive strategies, particu-

larly localized statin delivery, may offer pathways to enhance healing around implants under adverse metabolic conditions.

## STATINS AND BONE MINERAL DENSITY

Observational and trial data suggest statins may modestly improve bone mineral density (BMD). A meta-analysis involving over 34,000 participants (from both cohort and randomized controlled studies) found statistically significant increases in BMD at the lumbar spine (SMD 0.15), total hip (SMD 0.22), and femoral neck (SMD 0.19), with similar effect sizes across subgroups and ethnicities (16).

Preclinical studies reinforce that statins increase bone volume in animal models and suppress experimentally induced bone loss (10). These anabolic effects derive from both enhancement of osteoblast function and suppression of osteoclast activity, combined with potential improvements in bone vascularization and reduced apoptosis of bone cells (7, 8).

Currently, randomized trials focusing specifically on fracture outcomes are limited. The increase in BMD is modest and may not directly translate into reduced fracture risk for all patients. Differences in statin lipophilicity, dosage, duration, and baseline bone health may influence effectiveness. In short, statins may support BMD to a certain degree, especially over time, but their role in fracture prevention, osteoporosis and dental implant management remains suggestive rather than definitive (17).

## OSTEOBLAST PATHWAYS: STATIN INDUCED BONE FORMATION

**Statins inhibit HMG-CoA reductase**, disrupting the mevalonate pathway and reducing synthesis of isoprenoids like farnesyl pyrophosphate and geranylgeranyl pyrophosphate molecules essential for prenylation of small GTPases such as Rho and Ras (18). This inhibition impairs the prenylation dependent activity of Rho associated kinase (ROCK), which may relieve suppression on osteogenic signaling. Pitavastatin increased BMP-2 and osteocalcin expression in human osteoblasts, effects nullified by geranylgeranyl pyrophosphate, suggesting Rho-kinase inhibition mediates this enhancement of bone formation (19).

**Lovastatin activates Ras and downstream cascades.** It stimulated tyrosine phosphorylation and activation of the PI3K catalytic machinery via membrane bound Ras in osteoblasts. PI3K then activated Akt and Erk1/2, both of which contributed to upregulation of BMP-2 expression and several osteogenic markers including alkaline phosphatase, type I collagen, and osteopontin (20)

**Simvastatin harnesses a multi-pathway approach.** It enhances osteoblast viability and differentiation through a membrane-bound Ras → Smad1 → Erk → BMP-2 signaling cascade. The activation of Smad1 (a BMP effector) and Erk underscores how multiple molecular conduits may converge to foster osteogenesis (6).

**Summarizing**, statins may foster bone formation by:

1. Inhibiting Rho/ROCK via impaired prenylation

2. Activating Ras-PI3K-Akt/Erk signaling
3. Engaging Smad-mediated BMP-2 induction

These strands coalesce in a coordinated promotion of osteoblast differentiation, matrix synthesis, and survival, though the degree to which this translates to clinical bone repair or density gains may vary.

## OSTEOCLAST PATHWAYS: STATIN INDUCED INHIBITION OF RESORPTION

**Statins modulate the OPG/RANKL/RANK axis**, a critical control point in osteoclastogenesis. They elevate OPG (a decoy receptor) and reduce RANKL expression in bone-cell culture systems, dampening osteoclast formation (21).

**They also suppress NF-κB activation**, essential for osteoclast differentiation. Simvastatin inhibited RANKL-induced activation of NF-κB by blocking IκBα phosphorylation and degradation in RAW 264.7 cells, directly restraining osteoclastogenesis (9).

**Atorvastatin further acts by interrupting inflammatory signaling.** In synoviocytes from rheumatoid arthritis patients, it suppressed TNF-α-induced p38 MAPK phosphorylation and reduced RANKL expression and TRAP-positive osteoclast formation effects reversed by mevalonate supplementation, underscoring the role of the mevalonate pathway (22).

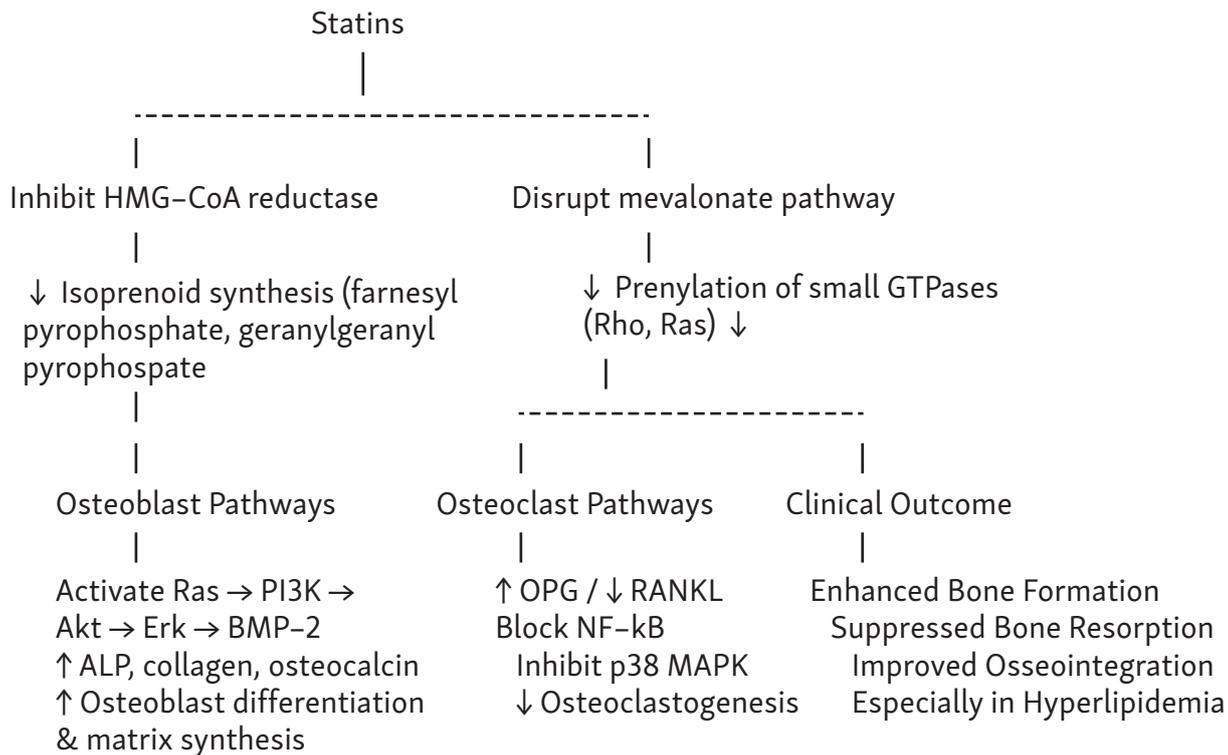
**In essence**, statins may attenuate bone resorption through multiple nodes:

1. Promoting OPG while suppressing RANKL
2. Blocking NF-κB activation required for osteoclastogenesis
3. Inhibiting p38 MAPK and related inflammatory triggers

## DISCUSSION

The experimental evidence presents a consistent mechanistic narrative that statins promote osteoblast differentiation via BMP-2 induction and Ras-PI3K-Akt-Erk signaling, while reducing osteoclastogenesis through modulation of the OPG/RANKL axis and suppression of NF-κB and inflammatory kinase activation (20). These complementary pathways could, in theory, favor net bone formation and improved osseointegration of dental implants (Graph 1). In hyperlipidemic animal models, statins also appear to counteract impaired osseointegration, suggesting possible translational relevance for implant dentistry.

Nevertheless, translation to clinical outcomes is limited by the modest magnitude of BMD changes observed in human studies, the absence of large scale trials targeting fracture endpoints, and variability in statin type, dose, and treatment duration. Additionally, obesity's partial but nondeterministic correlation with hypercholesterolemia reflects a broader metabolic milieu influencing both bone and cardiovascular health, making causality difficult to isolate. Future research should address whether targeted delivery systems, patient stratification by metabolic status, and longer treatment durations can reveal more



**Fig. 1** Effect of statins on bone remodeling and implant osseointegration.

substantial skeletal benefits. The convergence of lipid metabolism, inflammation, and bone remodeling pathways positions statins as promising, but as yet incompletely validated, agents in skeletal health management.

Future studies should optimize delivery vehicles that provide controlled, site specific release to the peri-implant region, minimizing systemic exposure and maximizing local BMP-2 induction.

Detailed *in vivo* and *ex vivo* studies are needed to confirm whether the peri-implant microenvironment, including exposure to oral microbiota, local inflammation, and masticatory forces, alters these signaling pathways.

Statins' modulation of the OPG/RANKL axis and suppression of NF-κB-mediated osteoclastogenesis should be investigated within peri-implant tissues, particularly under conditions of early implant loading or in patients with a history of periodontitis.

Given the links between obesity, dyslipidemia, and altered bone metabolism (23, 24), clinical trials should stratify patients by metabolic profile to determine whether hyperlipidemic or insulin resistant individuals derive greater peri-implant benefits from statin therapy.

Statin release from implant surfaces or coatings could be combined with micro/nano-textured titanium or bioactive ceramic surfaces to achieve additive or synergistic effects on osseointegration.

Most existing studies assess short term bone implant contact and early stability. Randomized controlled trials with extended follow up should evaluate whether statin based interventions reduce late implant loss, particularly in medically compromised patients.

A trial should explore whether statin therapy interacts with microbiota mediated metabolic and inflammatory pathways to influence peri-implant bone maintenance.

The limitations in future research design could be the heterogeneity of study models, the dosage and delivery methods, the duration of the follow up, the small sample sizes, the population diversity, the interaction with other medications and conditions (such as osteoporosis and diabetes mellitus), the molecular mechanism clarity and study design biases. Addressing these limitations will be crucial for using statins in dental implantology within the clinic, and for the design of future research. This approach will also maximize meaningful, translatable outcomes in evidence for these medications.

## CONCLUSION

Statins appear to be at least potentially bone friendly. They appear to enhance osteoblast activity, inhibit osteoclastogenesis, potentially provide some benefit in bone density along with the potential benefit in implant healing in the presence of hyperlipidemia. There is a relationship between obesity and hypercholesterolemia, yet non linearly, resembling the global form of metabolic interrelation involving obesity and hyperlipidemia. Therefore, this review will call for further clinical investigation, however, given what is presented in terms of mechanistic and pre-clinical evidence it is possible that the interrelated matrix discussed here deserves further examination.

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